

Research Article

Synthesis and *In Vitro* Evaluation of Novel Thieno [2,3 D] Pyrimidines As Antiproliferative Agents

Suparna S. De and Manisha S. Phoujdar

Sinhgad College of Pharmacy, Pune, 411041, Maharashtra, India.

ABSTRACT

The present study was undertaken to design and develop series of potential, selective antiproliferative-anticancer condensed pyrimidines, bioisosteric with gefitinib. Seven compounds were synthesized in three steps, structurally characterized and evaluated for their antiproliferative activity on HT29, A549 and HeLa cell lines which are found to overexpress EGFR. Compounds 1_h and 2_f showed good activity against HT29 cell line whereas compounds 1_i and 6_i showed good activity against HeLa and A549 cell lines respectively amongst the group.

Keywords: Condensed pyrimidines, antiproliferative, bioisosteric, HT29, A549, HeLa.

1. INTRODUCTION

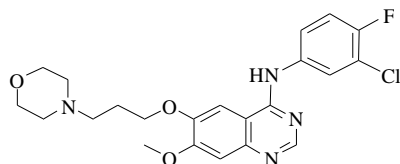
Carcinogenesis or malignant transformation of normal cells results in uncontrolled proliferation of some cells, as well as their metastasis to secondary foci. These undifferentiated extra cells form a mass called as tumour which could be malignant or benign. Currently anticancer agents possessing DNA cleaving, antimetabolic, antimitotic, topoisomerase inhibitory or signal transduction inhibitory properties are used as effective drugs, in the chemotherapy of cancer¹.

Several receptor tyrosine kinases² are known to be activated in the cancer cells and to drive tumour growth, angiogenesis, progression and metastasis. Inhibiting tyrosine kinase activity therefore represents a rational approach to cancer therapy³.

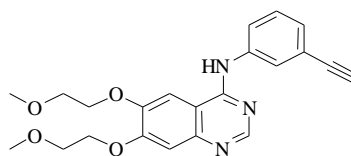
A variety of small molecules exhibiting selective receptor tyrosine kinase inhibitory activities have been reported as potential therapeutic agents to control tumour growth. Notable amongst them is the class of 2-*H*-4-anilinoquinazolines, suitably

substituted at the 5 and 6 positions. Noted antiproliferative drugs Gefitinib (Iressa)^{4,5}, Erlotinib (Tarceva)⁶, Lapatinib⁷, Canertinib⁸ etc., fall in this class (Fig.1).

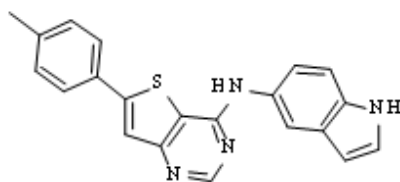
The bioisosterism between benzene and thiophene is well documented⁹. On these lines Munchhof *et al.*,¹⁰ have reported the design, synthesis and VEGFR-2 kinase inhibitory activity of some thieno[3,2-*b*]pyrimidines **3** (Fig.1). The potential of thienopyrimidines as antiproliferative anticancer activity is yet to be fully explored. On this basis, we undertook the exploration in the bioisosteric 4-anilino-2*H*-5, 6-disubstituted thieno[2,3-*d*]pyrimidines. Total seven compounds thus synthesized in good yields have been characterized structurally, and when evaluated on HT 29 (Adenocarcinoma)¹¹, A549 (Lung Carcinoma)¹² and HeLa (Cervix Cancer)¹³ cell lines for antiproliferative activity, have shown to have antiproliferative activity. These compounds have also been found non-cytotoxic to normal cells when tested on 3T3 normal mouse cell line.



1. Gefitinib



2. Erlotinib



3. Thieno[3,2-*b*]pyrimidines
Fig. 1: Structures of some antiproliferative drugs

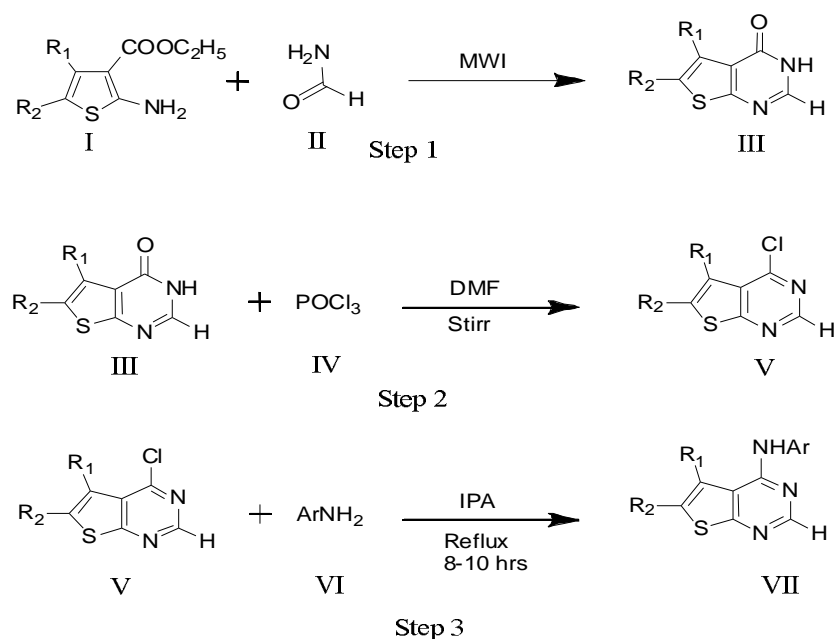
2. MATERIALS AND METHODS

The design of the novel condensed 2*H*-pyrimidin-4-amines has been done on the basis of the knowledge of antiproliferative activity associated with the 2*H*-4-anilinoquinazoline pharmacophore and its SAR [14]. On this basis a series of compounds, comprising of suitably substituted 2*H*-4-substituted-anilinothieno[2,3-*d*]pyrimidines (**1_{f,i}**, **2_{f,i}** and **6_i**) were designed, for the synthesis and biological evaluation against various carcinoma cell lines (HT29 cell line, A549 cell line and HeLa cell line).

2.1 Chemistry

The starting materials, *o*-aminocarbonyl substrates were synthesized in house^{15,16}. Their cyclocondensation with formamide in microwave, subsequent chlorination with POCl₃ and nucleophilic displacement reactions with various substituted anilines, to give a series of condensed 2*H*-pyrimidin-4-amines was done conventionally in three steps¹⁷.

A general approach to synthesize the designed compounds (**1_{f,i}**, **2_{f,i}** and **6_i**) is shown in Fig.2.



Where,

R₁=R₂=(CH₂)₄, R₁=R₂= CH₃, R₁=CH₃ R₂= COOEt,
 Ar = 3-ClC₆H₄, 3-CH₃C₆H₄, 4-CH₃C₆H₄, 3-BrC₆H₄

Fig. 2: Steps for synthesis of compounds (1_{f,i}**, **2_{f,i}** and **6_i**)**

The *o*-aminoesters (I) (0.02 mole) and formamide (II) (15 ml) were taken in 100 ml round bottom flask fitted with reflux condenser and irradiated in MWI at 40W for 25 min. The progress of the reaction was monitored by TLC. On completion, the reaction mixture was cooled to room temperature and poured into ice water. The solid obtained was filtered at suction, washed with chilled water and dried. The crude product (III) was recrystallised from methanol-dimethylformamide. This product (III) was then mixed with 40ml dimethyl formamide (DMF) and phosphorus oxychloride (IV) (10-12ml). The mixture was stirred for 1 hour at 0-5 °C and then stirring was continued at R.T for 8 hours. The progress of reaction was monitored by TLC. On completion of the reaction, excess of POCl₃ was distilled off under vacuum. The residue was then poured on crushed ice, neutralized by aq. NaHCO₃ and allowed to stand for one hour. The solid separated (V) was filtered by suction and washed with chilled water, dried and recrystallized from *n*-hexane. Thereafter, a solution of substituted aniline (VI) (0.02 mole) in isopropyl alcohol (20 ml) was added into the RBF containing compound V and the reaction mixture was refluxed conventionally for 8-10 hrs. The reaction mixture was cooled overnight. The solid separated (VII) was filtered at suction, washed with chilled water and dried. The crude product was recrystallised from *i*-propanol. The synthesized compounds were characterized by melting point, I.R, NMR and Mass. The list of synthesized compounds has been mentioned in Table 1.

2.1.1 Physical & spectral data for some representative compounds synthesized

N-(*m*-tolyl)-5,6,7,8-tetrahydrobenzo[4,5]thieno[2,3-*d*]pyrimidin-4-amine (1₇)

Melting Point: 128-130 °C

IR (KBr) cm⁻¹: 3456(ν_{NH}), 2921(ν_{C-H}), 1611(ν_{C=N}), 2857(ν_{CH3}).

¹H NMR(CDCl₃): 1.9 (4H, m, CH₂ at 6 and 7), 2.8 (2H, t, CH₂ at 8), 3.0 (2H, t, CH₂ at 5), 2.38 (3H, s, Ar-CH₃ at 3'), 6.9 (1H, d, Ar-H at 4'), 7.2 (1H, t, Ar-H at 5'), 7.44-7.48 (2H,d, Ar-H at 2' and 6'), 8.48 (1H,s, CH at 2)

N-(3-chlorophenyl)-5,6,7,8-tetrahydrobenzo[4,5]thieno[2,3-*d*]pyrimidin-4-amine (1₈)

Melting Point: 130-135 °C

IR (KBr) cm⁻¹: 3450(ν_{NH}), 2921(ν_{C-H}), 1600(ν_{C=N}), 764(ν_{C-Cl})

¹H NMR(CDCl₃): 1.89-1.98 (4H, m, CH₂ at 6 and 7), 2.8 (2H, t, CH₂ at 8), 2.9-3.0 (2H, t, CH₂ at 5), 4.0 (1H, NH at 4), 7.0 (2H, Ar-H at 2' and 4'), 7.2 (1H, t, Ar-H at 5'), 7.4 (1H, d, Ar-H at 6'), 8.48 (1H,s, CH at 2)

MS m/z. 316 (M⁺), 317 (M⁺¹), 318 (M+2), 288, 261, 189

2.2 Biological activity

The anti-proliferative potential of the compounds were tested on three different cell lines by MTT assay method.

The cell lines used were permanent cell lines obtained from National Centre for Cell Sciences (NCCS, Pune, India) and had origin from different cancers in humans. These were used as prototypes to find the potential of the compounds on different types of tumours. These cells have been characterized with the overexpression of EGFR¹⁸⁻²⁰.

2.2.1 Test Procedure

The HeLa, HT-29 and A-549 cell line were subcultured in a fresh tissue culture flask in DMEM (Dulbecco's Minimum Essential Medium) with 10% FBS (foetal bovine serum) and antibiotics, incubated in a 5% CO₂ incubator at 37°C. The cells were treated with appropriate concentration of each compound in triplicate with a control without any compound. The plate was incubated in a CO₂ incubator for 72 hr. and then studied under the inverted microscope and the results were recorded. The cell morphology in each well was compared with that of the control to analyze the results. The cells were incubated with the test sample in appropriate concentrations (10, 25, 50, 100 and 200µg/ml). The results were observed after 72 hr. under inverted microscope. A control cell lines was also maintained for each set of tests, which had only the respective cell line and the growth promoting media without any test compound. The test was performed in 2 sets.

Concentrations of compounds showing 50% growth inhibition in cells were calculated as IC₅₀ value.

3. RESULTS AND DISCUSSION

The activity comparison data of the synthesized compounds with the standard drugs (Gefitinib and Cyclophosphamide/CYPH) have been presented in Table 2. Data shows that all 7 compounds showed good inhibitory activity against three cell lines.

4. CONCLUSION

In present work, synthesis of a series of condensed 4-substituted anilino pyrimidines, unsubstituted at the position 2 of the pyrimidine ring has been undertaken. The moieties condensed to pyrimidine ring comprise of diverse structures like cyclohexane, dimethylthiophene and ethyl 3-methylthiophene-2-carboxylate. It was decided to try out these changes with respect to the ring attached to the pyrimidine ring, in order to obtain optimal activity. The starting materials, *o*-aminocarbonyl substrates were synthesized in house. Their cyclocondensation with formamide in microwave, subsequent chlorination with POCl_3 and nucleophilic displacement reactions with various substituted anilines was done conventionally. All the 7 compounds were found to have antiproliferative activity against the three cell lines. Amongst the group, compounds **1_h** and **2_f** have shown best activity against HT29 cell line whereas compounds **1_i** and **6_i** have shown best activity against HeLa and A549 cell lines respectively.

Conflict of interest statement

We declare that no benefits in any form have been received or will be received from a commercial party related directly or indirectly to the subject of this article. We also declare that we have no conflict of interest in connection with this paper, other than any noted in the covering letter to the editor.

REFERENCES

1. Foye WO, Lemke TL and Williams DA. Cancer and Chemotherapy. In: Lemke TL (ed) Foye's principles of medicinal chemistry. 5th edn. Lippincott Williams and Willikins, Baltimore. 2005; 924-5.
2. Manning G, Whyte DB, Martinez R and Hunter T, Sudarsanam S. The Protein Kinase Complement of the Human Genome. *Science*. 2002;298 (5600):1912-34.
3. Blume-Jensen P and Hunter T. Oncogenic kinase signalling. *Nature*. 2001;411(6835): 355-65.
4. Arteaga CL and Johnson DH. Tyrosine kinase inhibitors-ZD1839 (Iressa). *Curr Opin Oncol*. 2001;13 (6):491-8.
5. Barlesi F, Tchouhadjian C, Doddoli C, Villani P, Greillier L, Kleisbauer JP, Thomas P and Astoul P. Gefitinib (ZD1839, Iressa) in non-small-cell lung cancer: a review of clinical trials from a daily practice perspective. *Fundam Clin Pharmacol*. 2005;19:385-93.
6. Moyer JD, Barbacci EG, Iwata KK, Arnold L, Boman B, Cunningham A, Diorio C, Doty J, Morin MJ, Moyer MP, Neveu M, Pollack VA, Pustilnik LR, Reynolds MM, Slaon D, Theleman A and Miller P. Induction of apoptosis and cell cycle arrest by CP-358,774, an inhibitor of epidermal growth factor receptor tyrosine kinase. *Cancer Res*. 1997;57(21):4838-48.
7. Rusnak DW, Lackey K, Affleck K, Wood ER, Alligood KJ, Rhodes N, Keith BR, Murray DM, Glennon K, Knight WB, Mullin RJ and Gilmer TM. The effects of the novel, reversible epidermal growth factor receptor/ErbB-2 tyrosine kinase inhibitor, GW2016, on the growth of human normal and tumor-derived cell lines in vitro and in vivo. *Mol Cancer Ther* 2001;1(2):85-94.
8. Smaill JB, Palmer BD, Rewcastle GW, Denny WA, McNamara DJ, Dobrusin EM, Bridges AJ, Zhou H, Showalter HDH, Winters RT, Leopold WR, Fry DW, Nelson JM, Slintak V, Elliot WL, Roberts BJ, Vincent PW and Patmore SJ. Tyrosine kinase inhibitors. 15. 4-(Phenylamino)quinazoline and 4-(phenylamino)pyrido[d]pyrimidine acrylamides as irreversible inhibitors of the ATP binding site of the epidermal growth factor receptor. *J Med Chem*. 1999;4 (10):1803-15.
9. Foye WO, Lemke TL and Williams DA. Drug Design and Relationship of Functional Groups to Pharmacologic Activity. In: Lemke TL (ed) Foye's principles of medicinal chemistry. 5th edn. Lippincott Williams and Willikins, Baltimore. 2005;60-4
10. Munchhof MJ, Beebe JS, Casavant JM, Cooper BA, Doty JL, Higdon RC, Hillerman SM, Soderstrom CI, Knauth EA, Marx MA, Rossi AMK, Sobolov SB and Sun J. Design and SAR of thienopyrimidine and thienopyridine inhibitors of VEGFR-2 kinase activity. *Bioorg Med Chem Lett*. 2004;14:21-4.
11. Calonghi N, Pagnotta E, Parolin C, Tognoli C, Boga C and Masotti L. 9-Hydroxystearic Acid Interferes with EGF Signalling in a Human Colon

- Adenocarcinoma. *Biochem Biophys Res Commun.* 2006;342:585-8.
12. Rho JK, Choi YJ, Lee JK, Ryou BY, Na II, Yang SH, Kim CH and Lee JC. Epithelial to mesenchymal transition derived from repeated exposure to gefitinib determines the sensitivity to EGFR inhibitors in A549, a non-small cell lung cancer cell line *Lung Cancer.* 2009;63(2):219-26.
 13. Mitsuhashi A, Tanaka N, Suzuka K, Matsui H, Seki K and Sekiya S. Detection of epidermal growth factor receptor mRNA in peripheral blood of cervical cancer patients. *Gynecologic Oncology.* 2003;89 (3):480-5.
 14. Traxler PM. Protein Tyrosine Kinase Inhibitors in Cancer Treatment. *Expert Opinion on Therapeutic Patents.* 1997;7(6):571-88.
 15. Gewald K. Heterocycles from CH-acidic nitriles. VII. 2 Amino thiophene from α -oxo mercaptans and methylene-active nitriles. *Chem Ber.* 1965;98:3571.
 16. Gewald K, Schinke E and Bottcher H. 2-amino-thiophene aus methylenaktiven nitrilen, carbonylverbindungen und schwefel. *ChemBer.* 1966;99:94-100.
 17. Dabholkar VV . Total Microwave Based Synthesis of Novel Condensed Pyrimidine Analogs of Gefitinib. M. Pharm. Dissertation, Pune, India, 2008.
 18. Mendelsohn J. Blockade of receptors for growth factors: an anticancer therapy. *Clin Cancer Res.* 2000;6:747-53.
 19. Hu G, Liu W, Mendelsohn J, Ellis LM, Radinsky R, Andreeff M and Deisseroth AB. Expression of Epidermal Growth Factor Receptor and Human Papillomavirus E6/E7 Proteins in Cervical Carcinoma Cells. *Journal of the National Cancer Institute.* 1997;89 (17):1271-6.
 20. Kobayashi S, Kondo S and Juni K. Permeability of peptides and proteins in human cultured alveolar A549 cell monolayer. *Pharm Res.* 1995;12:1115-9.